

Study of Metallo-beta lactamase producing *Pseudomonas aeruginosa* in Pravara Rural Hospital

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Abstract

Carbapenem resistance is an emerging threat in the infections caused by gram negative pathogens. Detection of Metallo-beta lactamases (MBL) producing Pseudomonas is critical in preventing their widespread dissemination. Study was aimed at detection of Metallo-beta-lactamase in clinical isolates of Ps. aeruginosa by phenotypic methods in the Department of Microbiology, Rural Medical College, Loni. Out of eighty clinical isolates fifty four were subjected for MBL production by three different methods (i) Imipenem-EDTA double disc synergy method(DDST) (ii) Imipenem-EDTA combined disc test (iii) EDTA disc potentiation test using four cephalosporins. The MBL production was found to be 66.7%. This presents a therapeutic challenge to the clinicians and calls for judicious use of antibiotics.

Key words: *Ps.aeruginosa, MBL detection, Carbapenem resistance*

Introduction

In the past few decades *Pseudomonas aeruginosa* has been recognised as a pathogen in variety of infections and also an emerging nosocomial pathogen. *Ps. aeruginosa* shows intrinsic resistance to a variety of antimicrobials, including Beta lactams. The emergence of the MBL's in *Pseudomonas* species is fast becoming a therapeutic challenge as these enzymes possess high hydrolytic activity that leads to degradation of higher cephalosporins. Moreover, treatment options are either not available or very expensive and may be toxic with poor outcomes.^[1]

In view of the above, this study was undertaken to screen metallo beta-lactamase (MBL) production amongst multidrug resistant *Pseudomonas aeruginosa*.

Material and Methods

The study was conducted in the Department of Microbiology, Rural Medical College, Loni in 2009-2010. A total of 80 consecutive isolates of *Pseudomonas aeruginosa* obtained from various samples of the patients admitted in Pravara Rural Hospital were included in the study. All isolates were non duplicate. The isolates were identified as *Ps. aeruginosa* by conventional methods.^[2] Routine antibiotic disc sensitivity testing was done with penicillin, ampicillin, gentamicin, amikacin, ceftazidime, cefepime, ceftotaxime, ciprofloxacin, chloramphenicol, cefuroxime, cefpodoxime, norfloxacin, nitrofurantoin, cephalixin, cephazolin, sparfloxacin and cotrimoxazole. All the strains showing resistance to multiple drugs were tested for Imipenem susceptibility. Imipenem resistant isolates were further screened for MBL production.^[3-6]

1) Imipenem-Ethylenediaminetetraacetic acid (EDTA) double disc synergy test: Test organisms were inoculated on the plates with Muller Hinton

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agar as recommended by the CLSI (Clinical laboratory standards institute). An imipenem (10 microgram) disc was placed 20mm centre to centre from blank disc containing 10 microL of 0.5M EDTA (750 microgram). Enhancement of the zone of inhibition in the area between imipenem and the EDTA discs in comparison with the zone of inhibition on the far side of the drug was interpreted as positive (Fig1).

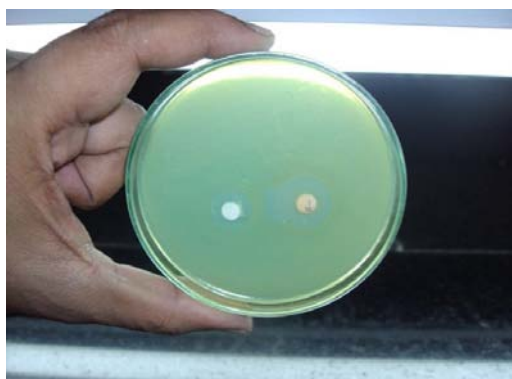


Fig 1: DDST positive

2) Imipenem (IPM)-EDTA combined disc test: Test organisms were inoculated on to the plates with Muller Hinton agar as recommended by the CLSI. Two 10 microgram imipenem discs were placed on the plates and appropriate amount of 10 microlitre of EDTA solution were added to one of them to obtain the desired conc. of 750 microgram. The inhibition zones were compared after 16-18 hrs of incubation at 35°C. In combined disc test, if the increase in inhibition zone was more than 7mm than the imipenem disc alone, it was considered as MBL positive. (Fig 2)

3) EDTA disc potentiation using cephalosporins: Test organisms were inoculated on Muller Hinton agar as described for the standard disc diffusion test. A filter paper, Whatman no.2 blank disc was placed and then following cephalosporins-Ceftazidime, Ceftizoxime, Cefotaxime and cefepime were placed



Fig 2: DDST Combined Positive

25mm center to center from blank disc. Ten microL of 0.5 M EDTA was added to the blank disc and plates were incubated at 35°C overnight. Enhancement of the zone of inhibition in the area between the EDTA disc and any one of the four cephalosporin disc in comparison with the zone of inhibition on the far side of the drug was interpreted as positive. ATCC 27853 *Pseudomonas aeruginosa* was used as negative control.^[3,4,5,6] Preparation of 0.5 M EDTA solution: 186.1g of disodium EDTA was dissolved in 1000 ml of D.W and pH adjusted to 8 by using NaOH. The mixture was sterilized by autoclaving.

3-2.jpg

Results

There were 80 isolates of *Ps. aeruginosa* in the study. Out of 80 isolates 54 Imipenem resistant isolates were screened for MBL production. 15 were isolated from pus, 13 from urine, 10 from blood, 5 from pleural fluid, 4 from vaginal swab, 3 from ICD catheter and 1 each from sputum, CSF, ear discharge & conjunctiva. All the strains exhibited resistance to most of the drugs (Table 1). Out of 54 Imipenem resistant isolates tested for MBL production 36 isolates exhibited more than 7mm zone size enhancement by the combined disc method whereas only 27 gave positive result by DDST and 6 isolates by EDTA disc potentiation method. Zone sizes were similar and reproducible when the procedure was repeated. Those isolates positive by DDST, were also positive by combined disc test for MBL production. The results with EDTA disc potentiation with four cephalosporins were not encouraging. Only six isolates showed zone enhancement towards EDTA disc in one or two antibiotics (Table 2).

Table 1: Antibiotic susceptibility testing showing resistance to various drugs

ANTIBIOTIC	RESISTANCE
Penicillin	100%
Ampicillin	92.86%
Amikacin	97.06%
Chloramphenicol	93.75%
Ceftazidime	96.43%
Ciprofloxacin	76.92%
Cephalexin	100%
Cephazolin	97.56%
Cephotaxime	100%
Cefuroxime	75%
Cefpodoxime	98%
Cefepime	92%
Gentamicin	85.71%
Norfoxacin	73.27%
Nitrofurantoin	100%
Nalidixic acid	100%
Cotrimaxazole	95.24%
Sparfloxacin	84%

Discussion

Pseudomonas aeruginosa is known to cause a variety of infections like urinary tract infection, wound infections, septicemia and device infections is now emerging as MBL producer and is spreading it in a horizontal way to members of Enterobacteriaceae. The injudicious or overuse of broad spectrum antibiotics has extended the incidence of MBL production in the community as well.^[7] These facts need to caution our clinicians and draw their attention towards the indiscrete use of antibiotics. In this study the imipenem resistance in *Ps. aeruginosa* was found to be 67.5% compared to 26% by A. Varaiya, 69% by Behera et al, 59.52% by S. Irfan, 30% by Gupta et al and only 8.05% by Agrawal et al.^[8,9,10,11]

In our study, by different methods, 66.7% (36/54 isolates) were found to be MBL producers. The remaining negative isolates may have other mechanism of resistance such as impermeability of outer membrane and or active efflux mechanism. A similar study carried out by Behera et al showed 48 isolates out of 64 as MBL

Table 2: Comparison of methods for detection of MBL *Ps. aeruginosa*.

Test	Total isolates tested (n=54)		
	DDST test	IMP-EDTA combined test	EDTA disc potentiation
ATCC 27853 <i>Ps. aeruginosa</i>	Negative	Negative	Negative
Isolates	27 (50%)	36 (66.7%)	06 (11.1%)

producers (76.19%) while 25 out of 25 i.e, 100% isolates were MBL producers in the study conducted by S. Irfan et al.^[8,9]

In our study we employed three methods for MBL detection. The incidence of MBL producing *Pseudomonas* by DDST was found to be 50% as compared to combined disc method (66.7%). In combined test it was easier to detect extension of zone more than 7 mm and therefore ease interpretations. The EDTA disc potentiation test with four cephalosporins was not found to be useful as it could detect only 6 isolates as MBL producers. The combined disc method proved to be more sensitive over DDST and EDTA disc potentiation methods in detection of MBL producers.^[12]

Conclusion

The study focuses on the resistance in *Ps. aeruginosa* towards Imipenem. This could be troublesome. Therefore there is need for implementation of surveillance studies in routine cases and careful selection of antibiotics in individual cases as well. The Imipenem-EDTA combined disc method is more convenient and useful for MBL screening. Emergence of MBL producing *Ps. aeruginosa* in clinical strains emphasizes the need in clinical practice to follow antibiotic restriction policies thereby avoiding

excess use of Carbapenems and other broad spectrum antibiotics. Since there are no standard guidelines for detection of MBL, the laboratories must evaluate the various methods for detection of MBL and use what is most suited. We believe that considering cost constraints, the combined disc test can be used as a convenient method.

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